Sun-shade variation in bamboo (Poaceae: Bambusoideae) leaves

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Abstract

The differential occurrence of fusoid cells in sun compared to shade leaves of three species of bamboo (Poaceae: Bambusoideae) was examined through leaf morphology and anatomy. Leaves grown in sun were consistently smaller and thicker and lacked fusoid cells, whereas leaves grown in shade were consistently larger, thinner, and had fusoid cells. These differences were also found between exterior and interior leaves within one individual of one of these three species. An infusion experiment using a fourth species indicated that light transmission increased dramatically as fusoid cells (or the spaces formed by their collapse) absorbed mineral oil. We propose that fusoid cells are a mechanism to trap and redistribute light more efficiently in shade leaves of bamboo and early-diverging grasses. This is the first report of leaf anatomical sun-shade variation in bamboo leaves, but we emphasize the preliminary nature of our observations and the need for further study.

Introduction

That sun and shade leaves differ in a general suite of characteristics is well documented; only representative references are cited here as this is not intended to be an exhaustive review. Leaves grown in direct sunlight are usually smaller and thicker with closer vein spacing compared to shaded leaves that are larger and thinner with more widely separated veins (e.g., Wylie 1949, 1951, Esau 1977, Cui et al. 1991, Ashton & Berlyn 1994, Yano & Terashima 2004, Terashima et al. 2006). Epidermal layers, especially the adaxial epidermis, are thicker in sun leaves than in shade leaves. Photosynthetic and light absorption characteristics, including mesophyll surface area, chloroplast distribution, resistance to CO$_2$ diffusion within the mesophyll, and rate of photosynthesis among others, also differ between sun and shade leaves (e.g., Nobel 1976, Boardman 1977, Lee et al. 1990, DeLucia et al. 1996, Terashima et al. 2001, Yano & Terashima 2004, Terashima et al. 2006).

Sun-shade leaf variation has been documented in a wide range of plant taxa, primarily angiosperms (e.g., references cited above, mainly for woody dicots) but also gymnosperms (e.g., Korstian 1925, Tucker & Emmingham 1977) and even...
pteridosperm fossils (Barbacka & van Konijnenburg-van Cittert 1998). Reports of sun-shade leaf variation within the grasses (Poaceae) confirm the same basic anatomical and physiological adaptations found in dicots (e.g., Slade 1970, Knapp & Gilliam 1985). Sun-shade differences within the canopy of a single individual have also been documented (Wylie 1949).

Hsiao et al. (1996) reported significant genetic differentiation between sun and shade subpopulations of *Yushania niitakayamensis* (Yushan cane, a temperate woody bamboo) in Taiwan. They also noted a difference in height between the sun and shade plants of this species, with sun plants generally reaching 1 m tall and shade plants growing up to 5 m tall, and cited Chen (1992) for additional information on variation in growth form in this species. This is the only report of sun-shade variation in bamboos that we found.

Among grasses, bamboos (Bambusoideae), the three early-diverging lineages (Anomochlooideae, Pharoideae, and Puelioideae), and *Streptogyna* P. Beauv. (*incertae sedis*) possess a characteristic leaf blade anatomy featuring in the mesophyll large, cigar-shaped, thin-walled, apparently empty cells known as fusoid cells and lobed or invaginated chlorenchyma cells known as arm cells (Brandis 1907, Page 1947, Soderstrom et al. 1987, Judziewicz et al. 1999, Grass Phylogeny Working Group 2001). As seen in cross section, the long axis of the fusoid cells is parallel to the epidermal layers, and typically a fusoid cell extends more or less perpendicularly from each side of a bundle (abutting the bundle sheath directly) into the middle of the mesophyll (Judziewicz et al. 1999, figs 14 & 15). Normally one layer of chlorenchyma cells is abaxial and one to three layers are adaxial to the fusoid cells (Page 1947, Judziewicz et al. 1999). As seen in leaf clearings, the fusoid cells are tightly packed in continuous longitudinal rows interrupted only by commissural veins (Judziewicz et al. 1999, fig. 16). The fusoid cells can occupy up to approximately 30% of the leaf blade volume (Clark 1991). Both Brandis (1907) and Page (1947) noted that the fusoid cells often (but not in all taxa) collapse, leaving large intercellular spaces.

Some taxa within Bambusoideae, however, are reported to lack fusoid cells. Among American woody bamboos, these include *Apoclada* McClure and *Filgueirasia* Guala (Guala 1995, 2003) and a few species of *Chusquea* Kunth (Clark 1986). The temperate woody genus *Phyllostachys* Sieb. & Zucc. is described as usually lacking fusoid cells (Watson & Dallwitz 1992 and onwards). Within Olyreae (herbaceous bamboos), *Ekmanochloa* A. Hitchc. and *Mniocloa pulchella* (Griseb.) Chase (Zuloaga et al. 1993), at least some species of *Raddiella* Swallen (Calderón & Soderstrom 1967) and *Parodiolyra ramosissima* (Trin.) Soders. & Zuloaga (Soderstrom & Ellis 1987) lack fusoid cells. At least some of these taxa occur in more open habitats (e.g., high altitude grassland species of *Chusquea*) but their lack of fusoid cells has never been investigated in terms of correlation with habitat.

Although fusoid cells are a prominent feature of the leaves of bamboo and early-diverging grasses, the possible function(s) of these cells remains unknown. Clark (1991) proposed that fusoid cells serve as reservoirs for CO$_2$ from photorespiration. We have not found any other hypotheses as to the function of fusoid cells to date.

Preliminary observations of two species of woody bamboos, *Yushania niitakayamensis* and *Chusquea culeou*, which occur in both shaded forest understories and adjacent clearings and grasslands (Lin 1978, Chen 1989, Pearson et al. 1994), revealed morphological and anatomical variation between sun and shade leaves, especially with respect to the presence or absence of fusoid cells. In our studies, we also included
Phyllostachys aurea (an Asian bamboo now widely cultivated) because of its putative lack of fusoid cells, and Bambusa longispiculata (another Asian bamboo often found in cultivation) because of its relatively large fusoid cells. We report our observations on sun-shade variation in bamboo leaves, with particular reference to the occurrence of fusoid cells, and propose a possible function for fusoid cells.

**Materials and Methods**

**Plant Material.** Yushania niitakayamensis and Chusquea culeou were collected in their natural habitats by March in Asia and by Clark in South America, respectively, as part of independent field work for systematic studies of each genus. Phyllostachys aurea and Bambusa longispiculata were cultivated in the R.W. Pohl Conservatory, Iowa State University (U.S.A.).

The following vouchers were used in this study:


**Paraffin Sections.** Paraffin-embedded sections were prepared from foliage leaf blades taken from herbarium specimens (*Y. niitakayamensis* and *C. culeou*) or from fresh leaves fixed directly in FAA (*P. aurea*). Whole leaf blades from herbarium specimens were soaked in Pohl’s solution (Pohl 1965) for at least seven days and then fixed for 24 hrs in FAA. Pieces approximately 1 cm² were taken from the mid-portions of the whole leaf blades and processed for embedding and sectioning. A total of 14 samples were prepared: two of *C. culeou* (one from each voucher), two of *P. aurea* (exterior and interior leaves from the same individual), and 10 of *Y. niitakayamensis* (two from *March 30* and one from each of the eight other vouchers).
Protocols for desilicification and dehydration followed Clark (1986). Leaf pieces were then bulk stained for one hour in 1% safranin in 1:1 ethanol:xylene, and rinsed for one hour each in two changes of xylene before infiltration for two days each in 2:1 xylene:paraffin, 1:2 xylene:paraffin, and 100% paraffin at 60°C. Samples were sectioned 10 µm thick on a rotary microtome, mounted on glass slides and stained in safranin (40 min) and fast green (4 min) using standard procedures (Sass 1958, Berlyn & Mikshe 1976).

**Hand Sections.** Hand sections were prepared from either fresh leaf material or leaves from herbarium specimens softened for a few minutes in Pohl’s solution (Pohl 1965). Sections were cut using a single-edged razor blade and immediately mounted and stained in a solution of lactophenol and aniline blue (Sass 1958). A few leaves per voucher were checked for consistency of fusoid cell development using this method.

**Leaf Infusion.** A leaf infusion process (Woolley 1971) involving mineral oil, which has a refractive index close to that of plant cell walls, was performed on fresh leaves of *Bambusa longispiculata* to determine the effect of fusoid cells on the transmission of light. For the infusion, whole fresh foliage leaf blades were coated with mineral oil, placed adaxial side down on microscope slides, and observed from the abaxial side under low magnification with a compound microscope at regular intervals (approximately every 20 minutes) for several hours.

### Results

**Morphology.** Sun-grown plants of both *Y. niitakayamensis* and *C. culeou* had shorter culms and smaller but relatively wider, lanceolate, foliage leaf blades compared with shade-grown plants (Fig. 1, Table 1). Likewise, foliage leaf blades of *P. aurea* grown in full sun were smaller, relatively wider and more lanceolate than those grown in the shade.

<table>
<thead>
<tr>
<th>Plant Height (m)</th>
<th>Leaf Shape</th>
<th>Mean Leaf Length (mm) (n = 10)</th>
<th>Mean Leaf Width (mm) (n = 10)</th>
<th>Mean Leaf Length: Width (n = 10)</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Yushania niitakayamensis</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Sun</td>
<td>&lt;0.8</td>
<td>lanceolate</td>
<td>33</td>
<td>4.0</td>
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<td>linear</td>
<td>104</td>
<td>7.2</td>
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<td><strong>Chusquea culeou</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Sun</td>
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<td>lanceolate</td>
<td>38</td>
<td>3.9</td>
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<tr>
<td>Shade</td>
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<td>linear</td>
<td>80</td>
<td>5.6</td>
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<tr>
<td><strong>Phyllostachys aurea</strong></td>
<td></td>
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<td></td>
<td></td>
</tr>
<tr>
<td>Sun</td>
<td>---</td>
<td>lanceolate and linear</td>
<td>107</td>
<td>17</td>
</tr>
<tr>
<td>Shade</td>
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<td></td>
</tr>
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</table>
Fig. 1. Leaf silhouettes of *Yushania niitakayamensis* shade (A), sun (B); *Chusquea culeou* shade (C), sun (D); and *Phyllostachys aurea* shade (E), sun (F).
Figs 2–7. Leaf cross sections showing anatomical differences between shade and sun leaves of *Yushania niitakayamensis* (Figs 2 & 3), *Chusquea culeou* (Figs 4 & 5) and *Phyllostachys aurea* (Figs 6 & 7) respectively.
Leaf Anatomy. Fusoid cells were present in the leaf blades of *Y. niitakayamensis* and *C. culeou* collected from shaded environments and from the lower, shaded leaves of the greenhouse-grown *P. aurea* plant (Figs 2, 4, 6; Table 2). In *C. culeou*, pairs of fusoid cells developed adjacent to every vein, while in *Y. niitakayamensis* and *P. aurea* the fusoid cells tended not to develop in the region nearest the margins. Between the marginal region where fusoid cells did not develop and the mid-leaf region where every fusoid cell developed, fusoid cells were usually either completely developed or completely undeveloped in a mosaic pattern, decreasing in occurrence rather than in size towards the margin. We could not determine from our preparations whether the fusoid cells died and remained intact or whether they uniformly collapsed to form spaces of the same size and shape as mature fusoid cells. Rarely, cells of the same size and in the same position as fusoid cells were observed, but these cells contained chloroplasts and had invaginated walls like the other mesophyll cells (Figs 8 & 9).

Figs 8 & 9. Cross sections of shade-grown leaves showing a large mesophyll cell in the position of a fusoid cell. Fig. 8. *Yushania niitakayamensis*. Fig. 9. *Phyllostachys aurea*. F = fusoid cell; LM = large mesophyll cell.
Fusoid cells were absent from the leaves of *Y. niitakayamensis* and *C. culeou* collected from fully sunlit sites and from the upper, exposed leaves of the *P. aurea* plant (Figs 3, 5, 7). No unusually large mesophyll cells were present in the positions of fusoid cells in these leaves. Arm cells in the adaxial-most layer remained asymmetrical but less strongly invaginated than when fusoid cells were present; the remaining mesophyll cells were symmetrically lobed (rosette cells or plicate mesophyll).

The average thickness of foliage leaf blades from plants of *Y. niitakayamensis* and *C. culeou* grown under sunny conditions were 62 and 79% greater, respectively, than those grown under shaded conditions (Table 2). Sun-exposed leaves of *P. aurea* were 30% thicker than shaded leaves from the same plant. Corresponding with this increased thickness was an increase from three to four or five layers of mesophyll cells. There were also increases in the thickness of the cuticle and both epidermal layers, most dramatically demonstrated in *C. culeou* (Figs 4 & 5).

The average distance between minor leaf veins was reduced in sun leaves compared with shade leaves of the same species: 8, 12 and 25% less for *Y. niitakayamensis*, *P. aurea*, and *C. culeou*, respectively (Table 2). Within each of the three species the number of major veins was more or less constant for both sun and shade leaves. However, the number of minor veins was consistently reduced in the sun leaves.

**Infusion experiment.** Fresh leaves of *Bambusa longispiculata* submerged in mineral oil changed from dark green to a notably lighter green as the oil infused into the fusoid cells or the intercellular spaces formed by their collapse (Figs. 10 & 11). When these fusoid cells (or spaces) were punctured at one end, a bubble of gas was released as the cell filled with oil. Concerted with this gas release and filling of each cell was an obvious further increase in the transmitted light. Over a period of three hours, mineral oil infused into about half of the fusoid cells (or intercellular spaces). Those remaining filled with gas were considerably darker (Fig. 11).

**Discussion**

**Differences between sun and shade leaves.** Consistent differences in leaf morphology and anatomy were observed between sun and shade plants or leaves in *Y. niitakayamensis*, *C. culeou* and *P. aurea*. The smaller leaf size, thicker leaf blades and closer vein spacing found in sun leaves are consistent with findings in other taxa exhibiting sun-shade variation, including the strong negative correlation between mean leaf length and light intensity index observed by Pearson et al. (1994) in *C. culeou*.

The increased thickness of the sun leaves of *Y. niitakayamensis* and *P. aurea* occurs despite the loss of fusoid cells but is correlated with the occurrence of an additional one or two layers of mesophyll cells as well as increased thicknesses of both epidermal layers and the cuticle. Individual layers of mesophyll cells are also thickened.

**Occurrence of fusoid cells.** The lack of fusoid cells in the sun leaves of the three species we examined is a dramatic modification of leaf anatomy, making sun and shade leaves appear to come from different species or genera, yet leaves with and without fusoid cells are even found on the same plant (*P. aurea*). The facultative occurrence of fusoid cells in bamboos has not previously been reported and should be considered when collections or anatomical studies of bamboos are made. This ontological correlation of fusoid cells with shade must be accounted for in any hypothesis of fusoid cell function.
The large mesophyll cells the size of fusoid cells, which were found in the position of fusoid cells in the zone where fusoid cell development is mosaic, may be intermediates having received a weak or incomplete developmental signal. This suggests that fusoid cells are homologous to mesophyll cells rather than to bundle sheath cells.

The pattern formed by collapsed fusoid cells is almost indistinguishable in clearings and cross sections from that formed by intact fusoid cells (Page 1947; P. Viana, pers. com.), so it is possible that collapse of fusoid cells is the usual condition at functional maturity of the leaf. We may in fact be observing a pattern of regular intercellular spaces formed by the normal collapse of the fusoid cells after they die, but the shape and volume of intact dead cells vs. the intercellular spaces created by their collapse appears to be equivalent and may well not affect the proposed functionality.

**Possible function of fusoid cells.** The suggestion that fusoid cells may serve as reservoirs for the retention of CO$_2$ from photorespiration (Clark 1991) is inconsistent with the differential occurrence of fusoid cells in shaded environments where photorespiration is less intense. We therefore reject this hypothesis.

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**Figs 10 & 11.** Leaf of *Bambusa longispiculata* infused with mineral oil, viewed from the abaxial side. Infused fusoid cells appear lighter, while uninfused fusoid cells appear darker. Fig. 10. Infusion for 6 minutes. Fig. 11. Infusion for 3 hours. AC = arm cells; F = fusoid cell; IF = infused fusoid cells; LV = leaf vein.
The difference in refractive index between air and cytoplasm subjects light entering a leaf to intercellular reflection, which can act to trap light within the leaf (Vogelmann 1989, 1993). DeLucia et al. (1996) concluded that intercellular reflectance contributes substantially to the absorption of light and that the increased proportion of spongy mesophyll in shade leaves may increase absorbance. The gas-filled array of fusoid cells (or comparably sized and shaped intercellular spaces) in bamboos may function in a similar way.

<p>| Table 2. Comparison of sun and shade leaf anatomy in <em>Y. niitakayamensis</em>, <em>C. culeou</em>, and <em>P. aurea</em>. |
|-------------------------------------------------------|-----------------------------------------------------|-----------------------------------------------------|</p>
<table>
<thead>
<tr>
<th>Fusoid cells</th>
<th>Mean Leaf Thickness (µm) (n = 10)</th>
<th>Mean Vein Spacing (µm) (n = 10)</th>
</tr>
</thead>
<tbody>
<tr>
<td><em>Yushania niitakayamensis</em></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Sun</td>
<td>absent</td>
<td>97</td>
</tr>
<tr>
<td>Shade</td>
<td>present</td>
<td>60</td>
</tr>
<tr>
<td><em>Chusquea culeou</em></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Sun</td>
<td>absent</td>
<td>138</td>
</tr>
<tr>
<td>Shade</td>
<td>present</td>
<td>77</td>
</tr>
<tr>
<td><em>Phyllostachys aurea</em></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Sun</td>
<td>absent</td>
<td>119</td>
</tr>
<tr>
<td>Shade</td>
<td>present</td>
<td>91</td>
</tr>
</tbody>
</table>

The observed increase in light transmission as fusoid cells (or intercellular spaces) were infiltrated with mineral oil was substantial (Figs 10 & 11), and indicates increased light absorbance in the native gas-filled state. Although not quantified, this observation is consistent with and lends support to the hypothesis that fusoid cells (or spaces formed by their collapse) contribute to light absorption through a mechanism of intra- or intercellular reflectance. This hypothesis is also supported by the ontological and phylogenetic correlations between fusoid cell occurrence and shade; under shaded conditions where photosynthesis is light-limited, enhanced light absorbance would be advantageous, but under sunny conditions where light is saturating or nearly saturating, enhanced absorbance would be less advantageous or even damaging.

**Conclusions.** Although this is a preliminary study, the results strongly suggest that fusoid cells function to trap and redistribute light in shade grown leaves of bamboos and, by extension, early-diverging grasses. Developmental studies of fusoid cells are needed to test the proposed homology of fusoid cells to mesophyll cells and also to test whether fusoid cells are indeed empty and dead at maturity and whether their collapse is routine or an artifact of processing. Field studies focused on the functional ecology of sun-shade variation in bamboos would also be of interest.
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